

FLUIDIC DEVICE WITH INTEGRATED CAPACITIVE MICROMACHINED ULTRASONIC TRANSDUCERS

5 Related Applications

This application claims priority to provisional application Serial No. 60/218,940 filed July 14, 2000.

Brief Description of the Invention

10 This invention relates to a fluidic device with integrated capacitive micromachined ultrasonic transducers (cMUTs), and more particularly to a fluidic device having microchannels with cMUTs fabricated in the walls of the channels.

Background of the Invention

15 The large investments in the microelectronics industry converted integrated circuits laboratories into machine shops where miniature electromechanical systems are designed and built. Electromechanical as well as electro-optical systems have been miniaturized and used in many different applications. In the same fashion, miniaturization is presently applied in the field of microfluidics. Microfluidics
20 technology provides the advantage of being able to perform chemical and biochemical reactions and/or separations with high throughput low volumes. Microfluidic systems employ microchannels in which chemical and biochemical materials are transported, mixed, separated and detected. The object is to take advantage of development in the silicon micromachining industry to develop laboratories on chips where fluids are
25 manipulated, transported and tested. Electric and optical fields form the backbone of

most of the methods used today in the transport and characterization of the fluids in channels.

Ultrasonic devices using piezoelectric materials have been successfully used for measurements of flow, physical properties and pressure of fluids and gases in many applications. Most of these devices are bulky, and they cannot be easily integrated to microfluidic systems for several reasons. With a few exceptions, piezoelectric materials are not compatible with other processing steps required for the fluidic chips. In addition, piezoelectric transducers for bulk wave excitation cannot be scaled down easily so as to fit in microfluidic channels without degrading their performance.

Summary and Objects of the Invention

Using recent developments in the field of ultrasonic sensors and actuators they can be integrated into microfluidic channels. The integration of ultrasonic transducers in small channels will enable many applications that have heretofore been the domain of large scale ultrasonic sensors and actuators. micromachined ultrasonic transducers (cMUTs) integrated in channels will be used in applications such as: fluid pumping, measurements of pressure, density, viscosity, flow rate and other fluidic properties.

Capacitive micromachined ultrasonic transducers (cMUTs) operating both in air and water are known and described in Patent Nos. 5,619,476, 5,870,351, 5,894,452. In both air and water, a Mason electrical equivalent circuit is used to represent the transducers and predict their behavior (W.P. Mason, *Electromechanical Transducers and Wave Filters* (Van Nostrand, New York, 1942)). These transducers are fabricated using standard IC processes and have been integrated with signal processing electronics to form an integrated system. In the article entitled "Highly Integrated 2-D Capacitive Micromachined Ultrasonic Transducers" appearing in IEEE Ultrasonic Symposium Proceedings pp. 1163-1666, 1999, S. Calmes et al. describe the fabrication of cMUTs with through wafer connections so that they can be flip-chip bonded to chips having signal processing electronics. The processing electronics can be implemented on the same silicon wafer avoiding the through wafer via structure. An example is provided in Figures 12 and 13. The dynamic range and bandwidth of cMUTs surpass their piezoelectric counterparts while being completely compatible with microfluidic chip fabrication processes.

cMUTs with dimensions of 100 μm or less are fabricated on the walls of the fluidic channels and operate in the 1-100 MHz frequency range. The cMUTs are surface micromachined to have a low surface profile, permitting undisturbed fluid flow. These transducers enable *in-situ* measurements of fluid flow, pressure, viscosity and temperature of the fluid in the channel. With their wide bandwidth, cMUTs can be used to implement resonators, time-of-flight measurements, and Doppler shift measurements in the fluid channel. It is also possible to excite traveling waves such as Stoneley waves at the fluid/channel wall interface to gently pump or mix fluids in the channel, in which case the cMUTs are used as actuators.

It is a general object of the present invention to provide fluidic channels having cMUTs fabricated in one wall of the channel for generating ultrasonic waves in said channel, and/or receiving ultrasonic waves.

Brief Description of the Drawings

Figure 1 is a schematic sectional diagram of a cMUT cell.

Figure 2 is a plan view of a cMUT array with five cells.

Figure 3 is a sectional view of a portion of a microfluidic channel including two cMUTs and signal processing electronics connected to the cMUTs by through wafer connections.

Figure 4 is a sectional view of the channel of Figure 2 taken along the line 3-3.

Figure 5 is a sectional view of a portion of a microfluidic channel employing a single cMUT for determining acoustic impedance of fluid in the channel.

Figure 6 is a sectional view of a portion of a microfluidic channel employing a single cMUT for determining fluid pressure.

Figure 7 is a sectional view of a portion of a microfluidic channel employing a single cMUT employing interdigitated cMUTs for generating Stoneley waves.

Figure 8 is a sectional view of a portion of a microfluidic channel employing two cMUTs for viscosity measurement.

Figure 9 is a top plan view of a mixer employing microfluidic channels and cMUT sensors.

Figure 10 is a sectional view taken along the line 10-10 of Figure 9.

Figure 11 is a plan view of mixer employing microfluidic channels and a cMUT mixer.

Figure 12 is a sectional view of a microfluidic channel including two cMUTs and on-wafer signal processing electronics.

Figure 13 is a sectional view taken along the line 13-13 of Figure 12.

5 Description of Preferred Embodiments

A cMUT cell is fabricated to form a structure similar to that of Fig. 1. The cell includes a substrate 11, such as silicon, and a membrane 12 such as silicon nitride supported by amorphous silicon 13. Amorphous silicon is used as a sacrificial layer that is partially removed by wet etching to form an evacuated cavity 14. A number of
10 cells 10 are fabricated on a silicon substrate to form a transducer 16, Figure 2. A detailed description of the methods for fabrication and operation of cMUTs is found in Patent Nos. 5,619,476, 5,870,351 and 5,894,452, incorporated herein in their entirety. In the illustrated embodiment, the gap thickness is determined by the amorphous silicon and can be quite small, which results in improved sensitivity because in
15 receive, one measures the change in capacity due to the motion of the membrane. Each cell is made of a vacuum-sealed, fully supported membrane with a diameter of 5-200 μm . For example, a 100 μm square transducer with individual cells 20 μm in diameter could be made with 25 such small membranes, Figure 2.

In microfluidic technology, the chemical or biochemical reactions and/or
20 separations take place in microchannels having dimensions in the range from 1 μm to 500 μm or more. Ultrasonic waves are ideal for measuring pressure, density, viscosity, flow rate and other properties of the fluids in the channels. Ultrasonic waves can also be used for fluid pumping. In accordance with the present invention, cMUTs are integrated into walls of the microchannels.

Referring to Figures 3 and 4, a microchannel 21 is shown in a section of a fluid
25 conduit or capillary 22. The microchannel can for example have dimensions of 1 μm to 500 μm or more, dependent upon the application. The channel can be formed by micromachining a groove 23 in the top plate 24 and suitably sealing it to a bottom substrate 26. The top 24 can be glass, silicon or the like, into which the groove is
30 machined, or it can be a polymeric material which can be machined or molded with the groove 23. In accordance with the present invention, the bottom substrate 26 is a semiconductor material such as silicon which is processed as described above to form integrated cMUTs such as cMUTs 27 and 28. The top surface of the cMUT is

substantially coextensive with the bottom wall of the channel, thereby minimizing the influence of the cMUTs on the fluid flow. The cMUTs can be connected to known excitation and detector electronics or processor 30 using through-wafer vias 29 and flip-chip bonding techniques such as those described by Oralkan (O.Oralkan, X.C. Jin, F.L. Degertekin and B.T. Khuri-Yakub, "Simulation and experimental characterization of a 2-D cMUT array element", *IEEE Trans. UFFC*, 46, pp. 1337-40, 1999).

Using the configuration of cMUTs shown in Figs. 3 and 4, the two cMUTs and the excitation and detection electronics are configured to alternately transmit and receive ultrasonic waves and to measure the times of flight of ultrasonic waves traveling along and opposite the direction of the flow. The difference in the time of flight in the two directions allows one to calculate the flow velocity of the fluid. With its wide bandwidth, the cMUT can easily separate ultrasonic pulses reflected up and down the fluidic channel. If the transmitting and receiving transducers are separated by 1 cm, and have the measurement ability to resolve 1/100 of a period at 100 MHz, it is possible to measure a flow velocity of 1 mm/sec. Several different frequencies can be used to provide different path lengths to enhance the accuracy of the measurement as depicted in the same figure. As commonly used in medical imaging, Doppler methods can also be utilized for flow measurement in the channel.

Other important physical parameters of the fluid in the channel can also be obtained *in-situ*. A pulse-echo measurement off the opposite wall gives the speed of sound in the fluid which is a measure of its stiffness divided by the density. Figure 5 is a sectional view showing a single cMUT 31 connected to a pulse echo processor 35. The processor excites the cMUT 31 which emits ultrasonic waves toward the opposite wall 32 and receiving the reflected waves. Multiple reflections between the walls of the channel can be used to set up resonance which can be used by the processor to determine the acoustic impedance, and hence the density and the viscosity of the fluid. The scattering from various structures in biological fluids, such as blood cells, can be detected using the cMUT in the pulse echo mode. This can be useful for both particle counting and Doppler shift.

The fluid pressure can be measured by a similar pulse-echo system monitoring the deformation of the channel. The fluid pressure will force the channel to deform in a predictable fashion, which in turn changes the path length of the reflected ultrasonic waves. In one embodiment, a compliant membrane 33 is fabricated on the wall

opposite the cMUT, Figure 6, with a vacuum-sealed gap 34 to reflect the ultrasonic wave in a pulse-echo measurement. The use of the vacuum-sealed gap will result in an absolute pressure measurement and a total reflection of the incident ultrasonic waves. The compliant membrane will have a large deflection for a given fluid pressure increasing the measurement sensitivity. For example, using a 0.4 μm thick 100 μm diameter silicon nitride membrane, deflections in the order of 1.5 \AA will be obtained for 1 Pa of fluid pressure. Using an ultrasonic time-of-flight (TOF) measurement with 1 ps resolution (off the shelf equipment can measure TOF down to 0.25 ps), one should obtain a pressure resolution of 5 Pa, assuming all other parameters, such as temperature, are calibrated out. As will be discussed later, an array of these compliant membranes and corresponding cMUTs can be placed along the channel to monitor the pressure drop due to the fluid flow and measure the fluidic resistance of the channel.

Since the dimensions of individual membranes forming the cMUTs are much smaller than the wavelength of the sound waves in the fluid, cMUTs generate significant evanescent fields in the fluid. In addition, at the edges, where the membranes are connected to the substrate, the motion of the cMUT membrane is coupled to the substrate. This combination results in an efficient excitation of propagating Stoneley waves at the fluid/substrate interface as shown in Fig. 6. Stoneley waves have an elliptical particle velocity field in the fluid that decays along the thickness of the channel. Hence, it is possible to move the fluid along the shallow channel by the traveling Stoneley waves which effectively turn the bottom surface of the channel into a distributed pump.

One can selectively excite Stoneley waves 36 while not coupling into the bulk waves in the channel by fabricating interdigitated cMUTs 32 on the wall of the fluidic channel as shown in Fig. 7. The mode selectivity is achieved by matching the spatial period of the cMUTs to the wavelength of the desired propagation mode. By applying in and out of phase signals to consecutive fingers, bulk wave radiation to the fluid can be avoided. By employing three spaced fingers or electrodes and applying 120° phase shifted signals, unidirectional fluid flow can be obtained. The traveling acoustic field in the channel has elliptical particle displacement fields that decay in the distance of $\lambda/2\pi$ from the excitation transducer surface, where λ is the wavelength of acoustic waves as shown in Fig. 7. For a water-like fluid, this will be around 24 μm for

Stoneley waves at 100 MHz. Hence, this frequency would be suitable for a typical channel height of 30 μm . At lower frequencies, the Stoneley wave will also couple to the top surface of the channel to generate plane wave-like modes traveling along the length of the channel. These modes will be useful in determining the flow rate of the fluid.

The Stoneley wave mode is evanescent in the case of a fluid/half-space structure and it will inherently provide more robust and repeatable sensors and actuators. These evanescent propagation modes will find many applications in measuring the properties of fluid and gas medium which flow in the microchannels.

Since the Stoneley waves are evanescent in the fluid, they propagate without damping if there is no loss in the fluid or solid substrate material. In a real fluid, the attenuation of these waves will be determined by the viscosity of the fluid. Hence, one can measure the fluid viscosity in a microfluidic channel by monitoring the amplitude of the Stoneley waves propagating a known distance in the channel. It has been shown that, for Lamb waves in thin plates, the insertion loss along a propagation path in dB is a linear function of fluid viscosity.

Another approach for viscosity measurement depends on the measurement of the fluidic resistance of the channel. The fluidic resistance of a channel with a rectangular cross-section and a length L is given by

$$R = \frac{\Delta P}{Q} = \frac{12\eta L}{wh^3}$$

where ΔP is the pressure drop in the channel in Pa, Q is the volume flow rate in m^3/s , w is the width, h is the height of the channel, and η is the viscosity of the fluid. Given the flow rate of the fluid and the pressure drop in the channel for a given length L , one can find the viscosity of the fluid for a given channel geometry. Combining the ultrasonic flow measurement with the pressure drop measured using an array of pressure sensors 31a and 31b as shown in Fig. 8, the viscosity can be monitored accurately. We note that the fluidic resistance expression is valid for a large w/h ratio, which would be valid in most cases. For microfluidic channels, the flow resistance, hence the pressure drops, may be significant even for small flow rates due to small dimensions. For example, for a 1 mm long water flow channel with 100 μm width and

30 μm height, the pressure drop will be ($R=8 \times 10^{12}$) 133 Pa (~ 1 Torr) for 1 $\mu\text{l}/\text{min}$ flow rate. If the pressure differences down to 5 Pa can be measured using the ultrasonic pulse-echo method, then a viscosity resolution of 0.07 centipoise can be achieved.

An example of the integration of cMUTs in the microchannels of a microfluidic device for fluid mixing and/or reaction is illustrated in Figures 9 and 10. The device includes a planar base 41 with integrated pairs of cMUTs 42 and 43, 44 and 45, and 46 and 47. Through wafer vias will carry electrical signals to the cMUTs. The pairs of cMUTs may be configured to generate Stoneley waves which would pump the fluid in the channels, or configured to measure the flow, or operated individually to sense pressure or other characteristics of the fluid in the channel. It is of course apparent that more cMUTs may be integrated to carry out the measurements discussed above.

In this example, a top glass wafer 51 is wet-etched to form input channels 52 and 53 and output channel 54. Fluid inlet and outlet ports 56 and 57 extend through the glass wafer to communicate with the channels. The glass wafer is suitably bonded to the planar base to form the microchannels over the cMUTs. The fluid flow through the input channels to the mixing chamber 58 and the reacted or mixed fluid flows through the outlet channel.

Figure 11 shows another embodiment of a fluid mixer. In this embodiment, the channels 52 and 53, Figure 9, merge smoothly into the channel 54. Parts in Figure 11 bear like reference numbers for like parts in Figure 9. The fluid which flows laminarily in the channels 52 and 53 travels as separate streams in the channel 54 and is mixed by action of one or both cMUTs 46 and 47.

As referred to above, the signal processing electronics can be connected to the cMUTs and carried on the surface of the wafer. Figures 12 and 13 show cMUTs 27 and 28 as in Figures 3 and 4 mounted in a microchannel 21. Signal processing and excitation integrated circuits 61 and 62 are mounted on the surface of the wafer 26 and connected along the surface of the wafer to the cMUTs rather than through vias.

The *in-situ* fluidic sensing and actuation schemes proposed for microfluidic channels enjoy the same advantages which has made the conventional, large-scale ultrasonic devices the popular choice for fluid measurements in industry. The high frequency cMUTs enable implementation of these techniques in microfluidic applications.

Especially in biological applications, it is critical to have fluidic sensors which do not interfere with the flow or affect the properties of the fluid. The microfluidic flow sensors based on dilution measurement of thermal, optical or ionic tracers require injection of heat, charge or light into the flow channel. Some other techniques

5 measure the drag force exerted on some specific structures inserted in the flow channel. Examples of these include capacitive or piezo-resistive measurement of the deflection of a cantilever placed in the flow channel. In most cases, these structures have to be fabricated separately and the flow channel is modified to fit the sensing structure disturbing the regular flow pattern. In contrast, the cMUTs are surface
10 micromachined to have a very low vertical profile and they will be an integral part of the channel wall. The ultrasonic sensors used for flow measurement do not require any thermal cycles or injection of tracers in the fluid flow, hence it is a non-intrusive technique.

Ultrasonic fluid pumping has inherent advantages due to its distributed-drive
15 mechanism as compared to the scaled down discrete pumps which require a drastic increase in the number of pumping stations and strength to keep up with the increased flow resistance in microfluidic channels. The cMUTs can operate at fairly low voltages to generate ultrasonic waves as compared to the pumps with direct electrostatic actuation. The fabrication of cMUTs are simple, all the micromachining
20 is performed on a single wafer using the standard semiconductor manufacturing techniques as opposed to electrostatically or magnetically actuated pumps with many hand-assembled moving parts. Also, the pumping is gentle; there are no thermal cycles or valve closures that could damage fragile biomolecules such as DNA. Furthermore, there are no restrictions on the type of fluid which may be pumped using
25 ultrasonic pumps. For example, hydrodynamic pumps cannot be used to pump conductive fluids.

The foregoing descriptions of specific embodiments of the present invention are presented for the purposes of illustration and description. They are not intended to be exhaustive or to limit the invention to the precise forms disclosed; obviously many
30 modifications and variations are possible in view of the above teachings. The embodiments were chosen and described in order to best explain the principles of the invention and its practical applications, to thereby enable others skilled in the art to best utilize the invention and various embodiments with various modifications as are

suited to the particular use contemplated. It is intended that the scope of the invention be defined by the following claims and their equivalents.

1. A method of determining the concentration of a substance in a sample, comprising the steps of: (a) measuring the absorbance of the sample at a wavelength of 254 nm; (b) measuring the absorbance of the sample at a wavelength of 280 nm; (c) measuring the absorbance of the sample at a wavelength of 300 nm; (d) measuring the absorbance of the sample at a wavelength of 320 nm; (e) measuring the absorbance of the sample at a wavelength of 340 nm; (f) measuring the absorbance of the sample at a wavelength of 360 nm; (g) measuring the absorbance of the sample at a wavelength of 380 nm; (h) measuring the absorbance of the sample at a wavelength of 400 nm; (i) measuring the absorbance of the sample at a wavelength of 420 nm; (j) measuring the absorbance of the sample at a wavelength of 440 nm; (k) measuring the absorbance of the sample at a wavelength of 460 nm; (l) measuring the absorbance of the sample at a wavelength of 480 nm; (m) measuring the absorbance of the sample at a wavelength of 500 nm; (n) measuring the absorbance of the sample at a wavelength of 520 nm; (o) measuring the absorbance of the sample at a wavelength of 540 nm; (p) measuring the absorbance of the sample at a wavelength of 560 nm; (q) measuring the absorbance of the sample at a wavelength of 580 nm; (r) measuring the absorbance of the sample at a wavelength of 600 nm; (s) measuring the absorbance of the sample at a wavelength of 620 nm; (t) measuring the absorbance of the sample at a wavelength of 640 nm; (u) measuring the absorbance of the sample at a wavelength of 660 nm; (v) measuring the absorbance of the sample at a wavelength of 680 nm; (w) measuring the absorbance of the sample at a wavelength of 700 nm; (x) measuring the absorbance of the sample at a wavelength of 720 nm; (y) measuring the absorbance of the sample at a wavelength of 740 nm; (z) measuring the absorbance of the sample at a wavelength of 760 nm; (aa) measuring the absorbance of the sample at a wavelength of 780 nm; (ab) measuring the absorbance of the sample at a wavelength of 800 nm; (ac) measuring the absorbance of the sample at a wavelength of 820 nm; (ad) measuring the absorbance of the sample at a wavelength of 840 nm; (ae) measuring the absorbance of the sample at a wavelength of 860 nm; (af) measuring the absorbance of the sample at a wavelength of 880 nm; (ag) measuring the absorbance of the sample at a wavelength of 900 nm; (ah) measuring the absorbance of the sample at a wavelength of 920 nm; (ai) measuring the absorbance of the sample at a wavelength of 940 nm; (aj) measuring the absorbance of the sample at a wavelength of 960 nm; (ak) measuring the absorbance of the sample at a wavelength of 980 nm; (al) measuring the absorbance of the sample at a wavelength of 1000 nm.